

NEOLIGNANS FROM *MEZILAURUS ITAUBA**

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Key Word Index—*Mezilaurus itauba*; Lauraceae; wood bark; dimeric lignoids; neolignans; benzofuranoid neolignans; bicyclo[3.2.1]octanoid neolignans.

Abstract—The wood bark of *Mezilaurus itauba* afforded in addition to seven known neolignans, three new compounds rel-(7*R*,8*R*,1'*S*,3'*S*)-Δ^{5',8'}-5'-methoxy-3,4-methylenedioxy-1',2',3',4'-tetrahydro-2',4'-dioxo-7,3',8,1'-neolignan, rel-(7*S*,8*S*,1'*S*,2'*S*,3'*R*,4'*S*)-Δ^{8'-2',4'}-dihydroxy-3,4-methylenedioxy-1',2',3',4',5',6'-hexahydro-5'-oxo-7,3',8,1'-neolignan and rel-(7*S*,8*S*)-Δ^{8'-6'}-hydroxy-5'-methoxy-3,4-methylenedioxy-7-*O*-2',8,3'-neolignan. The latter compound has been detected previously in *Aniba terminalis*. The structures were elucidated by spectroscopic methods and comparison with related compounds.

INTRODUCTION

Mezilaurus is a genus of the Lauraceae; comprising nine mostly arboreal species distributed mainly in the Amazon basin [1]. Only *M. syndra* has been investigated chemically and its trunk wood contains benzyl-isoquinoline alkaloids and γ -lactones [2].

The present paper describes the results obtained in a study of *M. itauba* (Meissn.) Taubert ex Mez., which grows in the Comisarias del Amazonas and Guaviare, Colombia; the plant is commonly known as itauba-stone wood, 'Japi-u-dlr' and 'Macmemorae' (dialects: Puinave and Witoto, respectively) [3]. The chloroform extract of the wood bark of *M. itauba* yielded seven benzofuranoid neolignans and three bicyclo[3.2.1]octanoid neolignans. The nomenclature and numbering of these neolignans follow the rules which were outlined in a recent review [4].

RESULTS AND DISCUSSION

Ten neolignans were isolated from *M. itauba*. Among these compounds 1b, 2a, 4a and 5a were reported earlier from *Aniba terminalis* [5, 6], neolignans of structural types 1b, 3a and 6a co-occur in *A. burchellii* [7, 8] and 7a was previously isolated from an *Aniba* species [9].

Compound 1a showed IR absorption at 3500 cm⁻¹ indicative of a hydroxyl group; confirmation of the presence of this functionality was provided by the UV spectrum ($\lambda_{\text{max}}^{\text{MeOH}}$ 325 nm; $\lambda_{\text{max}}^{\text{MeOH} + \text{NaOMe}}$ 344 nm). The complex UV spectrum of 1a, is indicative of a highly aromatic structure. Particularly informative was the ¹H NMR spectrum measured in carbon tetrachloride which showed a hydroxyl signal as a broad peak at δ 5.51, three aromatic protons as a multiplet at δ 6.68–6.90 and one aromatic proton singlet at δ 6.42. The AMX₃ system was analysed by ¹H NMR; (A): δ 4.93 (*d*, *J* = 8.0 Hz); (M):

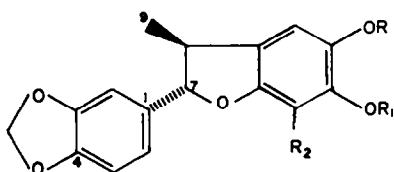
δ 3.10–3.50 (*m*); (X₃): δ 1.35 (*d*, *J* = 7.0 Hz). A *trans*-7,8-dihydrobenzofuran structure of 1a, was suggested by correlation with representatives of established structure [7]. An allyl group is also evident from the ¹H NMR spectrum which also exhibited signals due to methylene (δ 3.38, *d*, *J* = 6.0 Hz) and vinyl (δ 5.60–6.20, *m*, =CH and δ 4.80–5.20, *m*, =CH₂) groups as well as signals due to methoxy (δ 3.83, *s*) and methylenedioxy (δ 5.90, *s*) groups. Additional structural evidence was gained by preparation of 1c. Convincing evidence for the structure of 1a was provided by the mass spectrum ([M]⁺ at *m/z* 340 and fragment at *m/z* 162). This compound has been previously detected in a mixture [5].

For compound 7b, C₂₀H₂₀O₅ ([M]⁺ *m/z* 340), [α]_D –7.31°, the formula, established by mass spectrometry and expanded by inspection of UV, IR and ¹H NMR spectra, showed that the two C₆–C₃ units in this neolignan are linked only by C–C bonds and suggest that the compound is a bicyclo-octanoid [4]. Mass spectra were very helpful in defining one of the C₆–C₃ units by the fragment at *m/z* 162 (47%) which corresponded to an [ArCH=CHMe]⁺ fragment. The constitution of the second C₆–C₃ unit was defined by spectroscopic methods. Compound 7b showed IR absorptions at 1770 and 1718 cm⁻¹ (>CO) corresponding to part of the five-membered and six-membered rings, respectively. The stereochemical assignments of 7b are corroborated by ¹H NMR spectral comparisons with a model compound [10].

Compound 8a is a crystalline compound, mp 158–160°, [α]_D –0.69°, [M]⁺ *m/z* 330 showing aromatic ring absorption at λ_{max} 289 nm (log ϵ 3.51) in the UV spectrum and carbonyl absorption at 1698 cm⁻¹ in the IR spectrum. Absorption at 3504–3378 cm⁻¹ is attributed to hydroxyl groups and bands at 1640, 985 and 921 cm⁻¹ indicate the presence of an allyl group. The aliphatic nature of the hydroxyl group was deduced by inspection of the ¹³C and ¹H NMR spectra, which in addition gave further structural information (Table 1). C-2' and C-4' are represented by doublets at δ 79.0 and δ 75.7 and the two carbinol protons resonate at δ 4.60–4.76 (*m*, H-2') and

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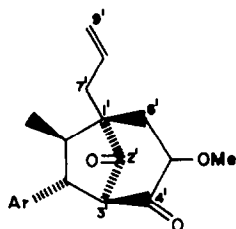
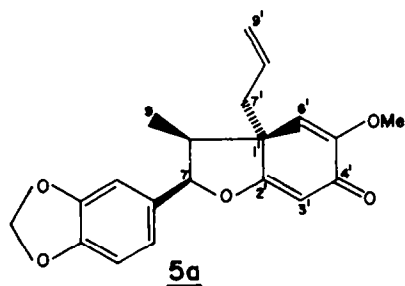
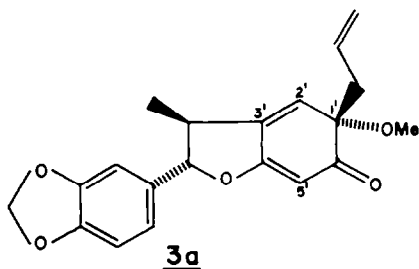
†To whom correspondence should be addressed.



1a R = Me, R₁ = H, R₂ = CH₂CH=CH₂

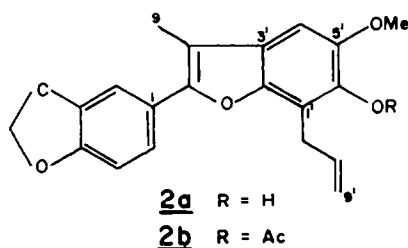
1b R = Me, R₁ = CH₂CH=CH₂, R₂ = H

1c R = Me, R₁ = Ac, R₂ = CH₂CH=CH₂



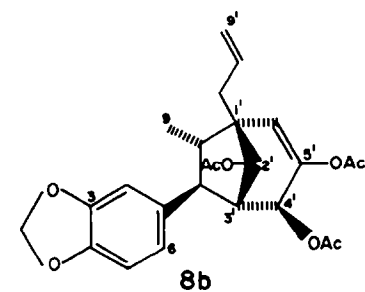
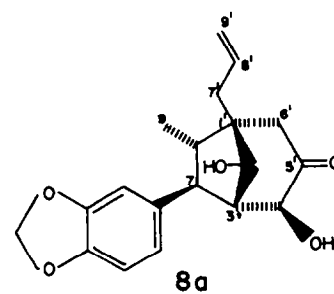
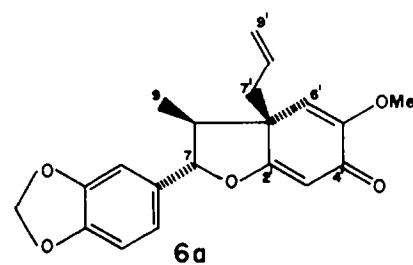
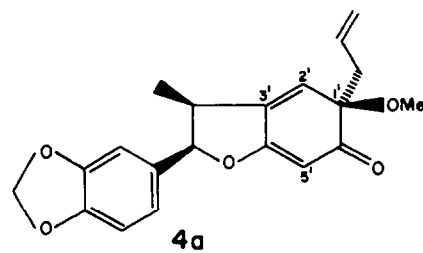
7a Ar = GUAIACYL

7b Ar = PIPERONYL



2a R = H

2b R = Ac



3.76–3.96 (*m*, H-4'). The aromatic ring is evident from two ¹³C oxygen bearing singlets at δ 148.7 and 147.6, three doublets at δ 108.1, 109.5 and 122.3 and a singlet at δ 140.4. This is confirmed by signals for three aromatic protons at δ 7.3 (*d*, J = 2.0 Hz, H-2), 6.78 (*d*, J = 6.0 Hz, H-5) and 6.78 (*dd*, J = 6.0 and 2.0 Hz) in the ¹H NMR spectrum which shows a signal for one methylenedioxy group at δ 5.96 (*s*). The remaining *sp*² ¹³C signals are ascribed to a carbonyl group at δ 210.5 and a monosubsti-

tuted double bond responsible for a doublet at δ 136.3 (C-8') and a triplet at 117.8 (C-9'). The substituent of the double bond is a methylene group responsible for a multiplet (δ 1.7–2.2), coupled to a vinyl proton that appears as a multiplet (δ 5.6–6.2, H-8') further coupled to two vinyl protons (δ 4.88–5.36, *m*, 2H-9'). In the ¹³C NMR spectrum C-7' appeared as a triplet at δ 38.6. The doublet at δ 0.87 (J = 7.0 Hz, 3H-9) indicate an Ar-7/Me-8-*trans*-relationship where the methyl is *endo* with

Table 1. ^{13}C and ^1H chemical shifts for **8a**

Assignment	^{13}C		^1H
	$\delta\text{Me}_2\text{CO}-d_6^*$	APT†	$\delta\text{Me}_2\text{CO}-d_6^\ddagger$
1	140.4	C	—
2	108.1	CH	7.30
3	148.7	C	—
4	147.6	C	—
5	109.5	CH	6.78
6	122.3	CH	6.78
7	55.2	CH	2.70
8	49.2	CH	2.20–2.60
9	12.3	Me	0.87
1'	51.7	C	—
2'	79.0	CH	4.60–4.76
3'	55.9	CH	2.20–2.60
4'	75.7	CH	3.76–3.96
5'	210.5	C	—
6'	43.4	CH_2	2.20–2.60
7'	38.6	CH_2	1.70–2.20
8'	136.3	CH	5.60–6.20
9'	117.8	CH_2	4.88–5.36
O_2CH_2	101.6	CH_2	5.96

* Measured at 20.0 MHz with TMS as internal reference.

† Information obtained from the APT.

‡ Measured at 60 MHz with TMS as internal reference.

respect to bicyclo [11]. The clearest evidence for the *endo* orientation of C-9 and C-6' was given by ^{13}C NMR signals at δ 12.3 (C-9) and δ 43.4 (C-6') corresponding to height values by the reciprocal γ -effect [12].

Acetylation transformed **8a** into the triacetate **8b**. A low resolution mass spectrum obtained at 30 eV, with significant peaks at m/z 456 (100%) $[\text{M}]^+$, 414 (5%), 355 (5%), 296 (3%) and 162 (59%), confirmed this reaction. This information, the strong IR band at 1748 cm^{-1} , the disappearance of the absorption at 1698 cm^{-1} present in **8a** and the number and chemical shifts of ^{13}C and ^1H NMR signals suggested the formation of acetate **8b** with enolization of **8a**. The acetate **8b** contains three acetoxy groups localized in 2' (δ 2.08, s, 3H; δ 20.86, q, 1C and δ 169.38, s, 1C), 4' (δ 2.13, s, 3H; δ 21.04, q, 1C and δ 169.79, s, 1C) and 5' (δ 2.16, s, 3H; δ 21.29, q, 1C and δ 170.39, s, 1C). Further inspection of the ^1H and ^{13}C NMR spectra of **8a** and **8b** is of particular relevance. In **8b** there are four olefinic protons (**8a** contains three olefinic protons) and the multiplet at δ 2.2–2.6 present in **8a** is absent. In the ^{13}C NMR spectrum there is disappearance of the signals at δ 43.4 (**8a**: t, 1C) and δ 210.50 (**8a**: s, 1C) and the appearance of a doublet at δ 121.44 (C-6') and a singlet at δ 147.93 (C-5'). Preliminary information about the number of directly attached protons to each carbon atom was obtained from APT (attached proton test) experiments (Table 1). Spectral comparison of **8a** and of its triacetate **8b** with canellin-C [11, 13] and other related compounds [9] indicates that **8a** and canellin-C to possess identical configurations.

EXPERIMENTAL

The plant material from the Bajo Guaviare, Comisaría del Guainía (Colombia), voucher Xiloteca de la Facultad de

Ingeniería Forestal, Universidad Distrital, Bogotá, Colombia, 424, was collected and identified by Drs. A. Roa and E. Acero.

Isolation of constituents. Wood bark (2 kg) was extracted with EtOH. The CHCl_3 soluble part (20 g) of the extract (76 g) was chromatographed on a silica S column (200 g). Fractions eluted with C_6H_6 were rechromatographed and further purified by prep. TLC (petrol– C_6H_6 and petrol– CHCl_3 with gradually increasing polarity) gave in order of elution, **2a** (16 mg), **1a** (35 mg), **1b** (143 mg) and sitosterol (31 mg). Fractions eluted with CHCl_3 were separated and purified by successive prep. TLC on silica gel HF₂₅₄. The developing solvents were different mixtures of petrol, hexane, EtOAc and Me_2CO and gave in order of elution, **5a** and **6a**-component mixture (79 mg), **7a** (23 mg), **3a** (66 mg), **4a** (17 mg), **7b** (18 mg) and **8a** (43 mg).

Identification of known compounds. The known compounds were identified by direct comparison of mp, $[\alpha]_D$ and spectral data with published data (**1b**, **2a**, **3a**, **4a**, **5a**, **6a** and **7a**). For these compounds only new data are included.

Rel-(7S,8S)- Δ^8 -6'-hydroxy-5'-methoxy-3,4-methylenedioxy-7-O-2',8,3'-neolignan (1a). Oil. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 224 sh (4.41), 293 (4.05), 308 sh (3.86), 325 (3.69), 345 sh (3.45); $\lambda_{\text{max}}^{\text{MeOH} + \text{MeONa}}$ nm: 223, 294, 309 sh, 344. IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 3500, 3075, 3003, 2956, 2922, 1636, 1610, 1504, 1490, 1466, 1443, 1388, 1337, 1245, 1216, 1118, 1096, 987, 930, 911, 871, 833, 799, 752. ^1H NMR (60 MHz, CCl_4): δ 6.68–6.90 (3H, m, H-2, H-5 and H-6), 4.93 (1H, d, J = 8.0 Hz, H-7), 3.10–3.50 (1H, m, H-8), 1.35 (3H, d, J = 7.0 Hz, H-9), 6.42 (1H, s, H-4'), 3.38 (2H, d, J = 6.0 Hz, H-7'), 5.60–6.20 (1H, m, H-8'), 4.80–5.20 (2H, m, H-9'), 5.90 (2H, s, OCH_2O), 5.51 (1H, s, OH-6'), 3.83 (3H, s, OMe-5'). MS 70 eV, m/z (rel. int.): 340 $[\text{M}]^+$ (100), 339 (12), 338 (44), 325 (16), 324 (10), 310 (4), 297 (13), 285 (10), 279 (8), 205 (20), 179 (4), 177 (5), 175 (5), 173 (7), 165 (7), 162 (4), 150 (3), 149 (20), 147 (9), 135 (17).

Rel-(7S,8S)- Δ^8 -6'-acetoxy-5'-methoxy-3,4-methylenedioxy-7-O-2',8,3'-neolignan (1c). (Ac_2O -pyridine, 24 hr room temp.). Oil. IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 3082, 2963, 2920, 2860, 1763, 1638, 1612, 1502, 1440, 1368, 1346, 1250, 1190, 1115, 1030, 990, 930, 889, 854, 755. ^1H NMR (60 MHz, CCl_4): δ 6.60–6.88 (3H, m, H-2, H-5 and H-6), 4.93 (1H, d, J = 8.0 Hz, H-7), 3.07–3.90 (1H, m, H-8), 1.36 (3H, d, J = 7.0 Hz, H-9), 6.46 (1H, s, H-4'), 3.20 (2H, d, J = 6.0 Hz, H-7'), 5.66–6.20 (1H, m, H-8'), 4.66–5.30 (2H, m, H-9'), 5.88 (2H, s, OCH_2O), 3.73 (3H, s, OMe-5'), 2.20 (3H, s, OAc-6'). MS 70 eV, m/z (rel. int.): 382 $[\text{M}]^+$ (36), 341 (14), 340 (100), 325 (18), 311 (2), 297 (9), 249 (3), 179 (4), 162 (2), 135 (15).

Rel-(7S,8S)- Δ^8 -6'-methoxy-3,4-methylenedioxy-7-O-3',8,4',1'-O-7'-neolignan (1b). $[\alpha]_D^{20} = +6.90^\circ$ (CHCl_3 ; c 4.2). ^{13}C NMR (20 MHz, CDCl_3): δ 144.28 (s, C-1), 106.37 (d, C-2), 147.82 (s, C-3), 147.33 (s, C-4), 107.84 (d, C-5), 119.44 (d, C-6), 92.59 (d, C-7), 45.64 (d, C-8), 18.12 (q, C-9), 134.76 (s, C-1), 97.28 (d, C-2'), 148.88 (s, C-3'), 122.25 (s, C-4'), 109.35 (d, C-5'), 153.31 (s, C-6'), 69.94 (t, C-7'), 133.41 (d, C-8'), 117.29 (t, C-9'), 100.76 (t, OCH_2O), 57.19 (q, OMe-6').

Rel-(7R,8R,1'S,3'S)- $\Delta^{5,8}$ -4-hydroxy-3,5'-dimethoxy-1',2',3',4'-tetrahydro-2',4'-dioxo-7,3',8,1'-neolignan (7a). Oil. $[\alpha]_D^{25} = -0.60^\circ$ (CHCl_3 ; c 0.83). Acetate of **7a** (Ac_2O -pyridine, 24 hr, room temp.). Oil. IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 3060, 2920, 2860, 1760, 1713, 1620, 1530, 1515, 1455, 1436, 1375, 1275, 1200, 1160, 1120, 1040, 1003, 920, 860. ^1H NMR (60 MHz, CCl_4): δ 6.50–6.90 (3H, m, H-2, H-5 and H-6), 3.81 (3H, s, OMe-3), 2.23 (3H, s, OAc-4), 2.68 (1H, d, J = 8.0 Hz, H-7), 1.83–2.45 (1H, m, H-8), 0.98 (3H, d, J = 7.0 Hz, H-9), 3.42 (1H, s, H-3'), 3.62 (3H, s, OMe-5'), 5.66 (1H, s, H-6'), 2.45–2.87 (2H, m, H-7'), 5.65–6.05 (1H, m, H-8'), 4.93–5.55 (2H, m, H-9'). MS 70 eV, m/z (rel. int.): 384 $[\text{M}]^+$ (6), 343 (24), 342 (100), 327 (5), 301 (19), 206 (1), 178 (7), 137 (12).

Rel-(7R,8R,1'S,3'S)- $\Delta^{5,8}$ -5'-methoxy-3,4-methylenedioxy-1',2',3',4'-tetrahydro-2',4'-dioxo-7,3',8,1'-neolignan (7b). Oil. $[\alpha]_D^{25} = -7.31^\circ$ (CHCl_3 ; c 0.20). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 236

(3.60), 289 (3.46). IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 3085, 2985, 2940, 2860, 1770, 1718, 1650, 1618, 1515, 1498, 1452, 1390, 1335, 1265, 1245, 1225, 1187, 1114, 1088, 1045, 1005, 938, 895, 872, 760. ^1H NMR (60 MHz, CCl_4): δ 6.42–6.79 (3H, m, H-2, H-5 and H-6), 2.47 (1H, d, $J = 7.3$ Hz, H-7), 1.77–2.20 (1H, m, H-8), 1.04 (3H, d, $J = 6.6$ Hz, H-9), 3.56 (1H, s, H-3'), 5.64 (1H, s, H-6'), 2.25–3.05 (2H, m, H-7'), 5.60–6.12 (1H, m, H-8'), 4.95–5.36 (2H, m, H-9'), 3.67 (3H, s, OMe-5'), 5.92 (2H, s, OCH_2O). MS 70 eV, m/z (rel. int.): 340 $[\text{M}]^+$ (100), 325 (6), 312 (2), 299 (27), 284 (10), 271 (27), 243 (5), 178 (5), 162 (47), 150 (4), 137 (6), 135 (25).

Rel-(7S,8S,1'S,2'S,3'R,4'S)- $\Delta^{5,8}$ -2',4'-dihydroxy-3,4-methylenedioxy-1',2',3',4',5',6'-hexahydro-5'-oxo-7,3',8,1'-neolignan (8a). Crystals, mp 158–160° (CCl_4). $[\alpha]_{\text{D}}^{25} = -0.69^\circ$ (Me_2CO ; c 2.15). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 237 (3.58), 289 (3.51). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3504, 3450, 3378, 3078, 3014, 2956, 2915, 1698, 1640, 1610, 1501, 1485, 1451, 1390, 1321, 1250, 1210, 1185, 1152, 1118, 1100, 1070, 1040, 1015, 985, 963, 938, 921, 892, 880, 795, 730. ^1H NMR (60 MHz, $\text{Me}_2\text{CO}-d_6$): see Table 1; ^{13}C NMR (20 MHz, $\text{Me}_2\text{CO}-d_6$) and APT experiment, see Table 1. MS 30 eV, m/z (rel. int.): 330 $[\text{M}]^+$ (100), 271 (6), 240 (15), 199 (3), 177 (7), 149 (3), 162 (13), 135 (4).

Rel-(7S,8S,1'S,2'S,3'R,4'S)- $\Delta^{5,8}$ -2',4',5'-triaceoxy-3,4-methylenedioxy-1',2',3',4'-tetrahydro-7,3',8,1'-neolignan (8b). (Ac_2O -pyridine, 120 hr room temp.). Oil. IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 3074, 3019, 2960, 2920, 2872, 1748 vs, 1668, 1641, 1610, 1502, 1486, 1442, 1368, 1220 s, 1035, 978, 923, 858, 810, 790, 748. ^1H NMR (60 MHz, CDCl_3): δ 6.70–6.95 (3H, m, H-2, H-5 and H-6), 2.10–2.80 (2H, m, H-7 and H-8), 0.99 (3H, d, $J = 7.0$ Hz, H-9), 5.60 (1H, br s, H-2'), 2.10–2.80 (1H, m, H-3'), 4.88–5.08 (1H, m, H-4'), 5.30 (1H, br s, H-6'), 1.70–2.10 (2H, m, H-7'), 5.55–6.10 (1H, m, H-8'), 4.88–5.55 (2H, m, H-9), 5.94 (2H, s, OCH_2O), 2.08 (3H, s, OAc-2'), 2.13 (3H, s, OAc-4'), 2.16 (3H, s, OAc-5'). ^{13}C NMR (20 MHz, CDCl_3): δ 134.20 (s, C-1), 108.10 (d, C-2), 146.10 (s, C-3), 143.84 (s, C-4), 108.20 (d, C-5), 125.97 (d, C-6), 53.02 (d, C-7), 50.83 (d, C-8), 12.75 (q, C-9), 50.65 (s, C-1'), 73.77 (d, C-2'), 53.02 (d, C-3'), 77.60 (s, C-4'), 147.93 (s, C-5'), 121.44 (d, C-6'), 36.10 (t, C-7'), 137.08 (d, C-8'), 117.76 (t, C-9'), 100.91 (t, OCH_2O), 20.86 (q, MeCO-2'), 21.04 (q, MeCO-4'), 21.29 (q, MeCO-5'), 169.38 (s, MeCO-2'), 169.79 (s, MeCO-4'), 170.39 (s, MeCO-5'). MS 30 eV, m/z (rel.

int.): 456 $[\text{M}]^+$ (100), 414 (5), 355 (5), 354 (9), 336 (9), 296 (3), 295 (11), 294 (31), 276 (8), 253 (9), 162 (59), 135 (7).

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